

# Whole exome sequencing as a first line test: Is there even a role for metabolic biochemists in the future?

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# Whole exome sequencing

No doubt this is the way forward for “difficult” patients.  
But:

- Still slow - months
- Too much data - difficult to interpret, especially in consanguineous families
- Still relatively expensive
- A sledge hammer to crack a nut?

At this stage, whole exome sequencing major impact on a small number of families. Some referrals trickling through for confirmatory tests.

Limited clinical exome sequencing  
for the diagnosis of inherited  
metabolic disorders  
or  
The end of metabolic biochemistry  
as we know it ....

# The problem

- There is no agreement on what constitutes an IMD or how many there are - ?? more than 550?
- Individually rare, cumulative incidence is about 1 in 1,500 to 1 in 5,000 live births.
- Considerable variation in clinical presentations
- **No first line biochemical test picks up all disorders.**
- Most biochemical tests done for purposes of exclusion
- Just one third of patients are diagnosed by the age of 1 year

RESEARCH

Open Access

## The diagnosis of inherited metabolic diseases by microarray gene expression profiling

Monica Arenas Hernandez<sup>1</sup>, Reiner Schulz<sup>2</sup>, Tracy Chaplin<sup>3</sup>, Bryan D Young<sup>5</sup>, David Perrett<sup>6</sup>, Michael P Champion<sup>7</sup>, Jan-Willem Taanman<sup>8</sup>, Anthony Fensom<sup>4</sup>, Anthony M Marinaki<sup>1\*</sup>

**Microarray gene expression profiling – identified the defective gene in 14/68 (21%) of cell lines. Frame shift mutations result in nonsense mediated decay and low mRNA levels**

Slow and expensive - need to establish fibroblast cell lines

# The solution

Limited exome sequencing for known genetic disorders would, in a single first line test, diagnose 80-90% of IMD cases.

**The concern:** raised by Metabolic Consultants at the Evelina, false positive results would add to diagnostic costs.

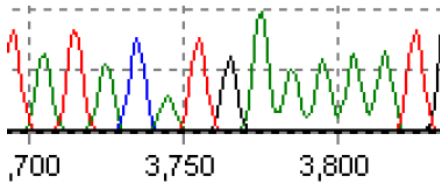
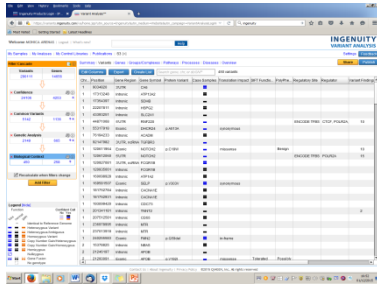
**The challenge:** find the genetic defect in 9 previously diagnosed patients.

**The catch:** we would be blinded to all clinical and demographic information.

# TruSight™ One Sequencing Panel (Illumina)

- Targets exonic regions (including promoter, UTR-5' and UTR-3')
- Genomic targets derived from:
  - Human Gene Mutation Database (HGMD Professional)
  - Online Mendelian Inheritance in Man (OMIM) catalog
  - GeneTests.org
  - Illumina TruSight panels
- Panel = 4813 genes of which 2761 genes are derived from the Human Gene Mutation Database
- High depth of coverage (>20x) for more than 95% of the targets for multiple samples sequenced on a MiSeq.

# Time lines



Bench work, 3 samples multiplexed  
3 days

Run time on MiSeq  
3 days

Ingenuity, web-based software for variant analysis  
1 day

**Preliminary diagnosis**

Sanger sequencing for confirmation  
1 week, or biochemical confirmation 2 days

**Confirmed diagnosis**



# Results

## One false negative

The diagnosis could not be made in one patient.

- After un-blinding: diagnosis of GSD IV
- Re-examination of data set: heterozygous c.476C>T p.P159L
- There is a rare/private variant in the 5'UTR c.37-36insC
- We cannot make the call on a single, although probably deleterious variant.

# The trouble with panels ....**the gene defect was filtered out in two patients**

## **Vanishing white matter disease**

- *EIF2B5* c.116-130del, p.39-L43 homozygous
- Eukaryotic translation initiation factor 2b, subunit 5, not a metabolic disorder as such.

## **Imlerslund Grasbeck syndrome (megaloblastic anaemia)**

- *CUBN* c.269C>G p.S897\* and c.796G>A p.E266K
- Cubilin is an intestinal transporter for uptake of IF-B12

# Red herrings: Imerlund Grasbeck syndrome

- *GALC* c.1514G>A, p.R515H heterozygous (Krabbe's disease)
- *BCKDHA* c.857-1G>C, p.A285P heterozygous (Maple syrup urine disease)
- *ABCD1* c.1816T>C p.S606P heterozygous (X-linked adrenoleukodystrophy) – multiple pseudogenes have this variant.

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|         |            |   |   |            |            |            |            |            |
|---------|------------|---|---|------------|------------|------------|------------|------------|
| abcd1g  | ACGTCCTG   | T | C | GGGTGGCGAG | AAGCAGAGAA | TCGGCATGGC | CCGCATGTTC | TACCACAGGT |
| abcd1p1 | ACGTCCTGCC |   |   | GGGTGACGAG | AAGCAGAGAA | TCGGCATGGC | CCGCATGTTC | TACCACAGGT |
| abcd1p2 | ACATCCTGCC |   |   | AGGTGGTGAG | AAGCAGAGAA | TCGGCATGGC | CCGCATGTTC | TACCACAGGT |
| abcd1p3 | ACGTCCCCGC |   |   | GGGTGGCGAG | AAGCAGAGAA | TCGGCATGGC | CCGCATGTTC | TACCACAGGT |
| abcd1p4 | ACGTCCTGCC |   |   | GGGTGGCAAG | AAGCAGAGAA | TCGGCATGGC | CTGCATGTTC | TACCACAGGT |
| abcd1p5 | ATGTCCTGCC |   |   | GGGTGGCGAG | AAGCAGAGAA | TCGGCATGGC | CCGCATGTTC | TACCACAGGT |

# OMIM search: megaloblastic anaemia

| OMIM Disorder   | Gene  |
|---|---|
| 219721 - CYSTIC FIBROSIS WITH HELICOBACTER PYLORI GASTRITIS, MEGALOBLASTIC ANEMIA, AND MENTAL RETARDATION | No gene reported  |
| #236270 - HOMOCYSTINURIA-MEGALOBLASTIC ANEMIA, cbIE COMPLEMENTATION TYPE; HMAE                            | MTRR No variants  |
| #249270 - THIAMINE-RESPONSIVE MEGALOBLASTIC ANEMIA SYNDROME; TRMA   | SLC19A2 No variants   |
| #250940 - HOMOCYSTINURIA-MEGALOBLASTIC ANEMIA, cbIG COMPLEMENTATION TYPE; HMAG                            | <b>MTR 11 variants all polymorphic.</b>                       |
| #261100 - MEGALOBLASTIC ANEMIA 1  | <b>CUBN 12 polys, 2 causative variants</b><br>AMN no variants |
| #275350 - TRANSCOBALAMIN II DEFICIENCY  | TCN2 No variants  |
| #615631 - ANEMIA, CONGENITAL DYSERYTHROPOIETIC, TYPE Ib; CDAN1B   | C15orf41 No variants  |
| #300322 - LESCH-NYHAN SYNDROME; LNS   | <b>HPRT1 no variants</b>                                      |
| #277400 - METHYLMALONIC ACIDURIA AND HOMOCYSTINURIA, cbIC TYPE  | <b>MMACHC 1 poly</b>  |
| #224120 - ANEMIA, CONGENITAL DYSERYTHROPOIETIC, TYPE Ia; CDAN1A   | CDAN1 No variants   |
| #236200 - HOMOCYSTINURIA DUE TO CYSTATHIONINE BETA-SYNTHASE DEFICIENCY                                    | CBS no variants   |

vic

# In six out of nine patients a definitive diagnosis was made

| Gene         | Mutation  | Disorder                                     |
|--------------|---|--|
| <i>GCDH</i>  | c.442G>A p.V148I<br>c.641C>T p.T214M<br>c.548A>G, p.N183S | Glutaric acidaemia type 1                    |
| <i>PEX1</i>  | c.2528G>A, p.G843D<br>homozygous                          | Peroxisome Biogenesis Disorder (Zellweger's) |
| <i>MMAB</i>  | c.556C>T, p.R186W<br>c.700C>T, p.234Q>*                   | Methylmalonic acidaemia, cblB TYPE           |
| <i>IVD</i>   | c.367 G>A p.G123R<br>Homozygous                           | Isovaleric acidaemia                         |
| <i>GLDC</i>  | c.2964G>A, p.R988Q<br>c.335-5A>G                          | Non-ketotic hyperglycinaemia                 |
| <i>PHKA2</i> | c.1005delT, p.F335fs*2<br>hemizygous                      | Glycogen storage disease Type IXa            |

# Non-ketotic hyperglycinaemia

|             |  |                                     |
|-------------|--|-------------------------------------|
| <b>GLDC</b> | <b>c.2964G&gt;A, p.R988Q</b><br><b>c.335-5A&gt;G</b> | <b>Non-ketotic hyperglycinaemia</b> |
|-------------|--|-------------------------------------|

## **c.2964G>A, p.R988Q**

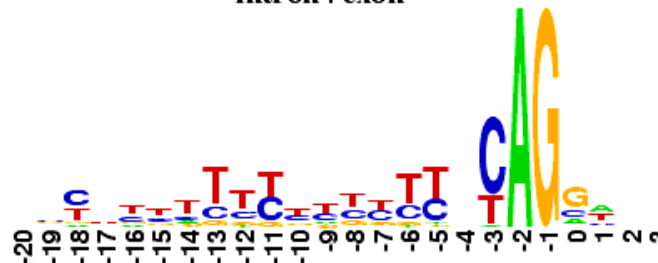
- Previously unreported variant
- PolyPhen prediction is that the amino acid substitution is **PROBABLY DAMAGING**

## **c.335-5A>G, splice site changed. Previously unreported variant**

Authentic 3'-splice site: intron 2-tttttcccaca**a**ttag/GTGAAA-exon3

Variant splice site: intron 2-tttttcccaca**g**/TTAGGTGAAA-exon3

intron | exon



# What we learned from the pilot study

A retrospective study using the TruSight One panel run on a MiSeq

- The diagnosis was made in 8 of 9 patients: organic acidaemias, peroxisomal, glycogen storage, cobalamin transport and a defect in eukaryotic translation initiation factor 2b, subunit 5 - not a metabolic disorder as such.
- Pseudogenes and heterozygous variants of unknown significance are a problem
- Focused gene panels do not reflect real world clinics

# What metabolic clinicians want -

A single test to diagnose the majority of disorders

- Fast – clinically relevant TAT
- Affordable/cost effective
- Low false-positive rate
- Confirmation of the defect by a second measure – metabolites, enzyme assay etc

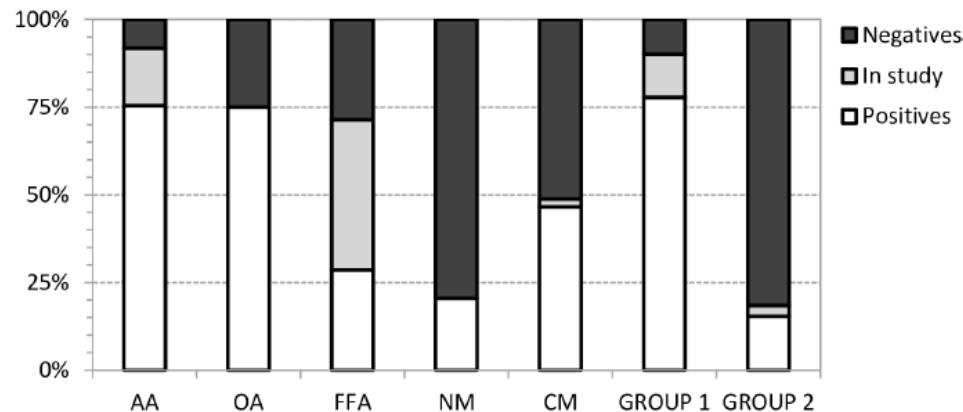


RESEARCH ARTICLE

# Targeted Next Generation Sequencing in Patients with Inborn Errors of Metabolism

Dèlia Yubero<sup>1</sup>, Núria Brandi<sup>2,3</sup>, Aida Ormazabal<sup>1,6</sup>, Àngels Garcia-Cazorla<sup>4,6</sup>, Belén Pérez-Dueñas<sup>4,6</sup>, Jaime Campistol<sup>4,6</sup>, Antonia Ribes<sup>5,6</sup>, Francesc Palau<sup>3,6</sup>, Rafael Artuch<sup>1,6</sup>, Judith Armstrong<sup>3,6</sup><sup>\*</sup>, Working Group<sup>1</sup>

## Global genetic results



**Fig 1. Global genetic results.** Genetic results (positive, under-study, and negative cases) shown as a percentages for each nosological group (aminoacidopathies (AA); organic acidurias (OA); free fatty acid oxidation defects (FFA); and neurometabolic (NM) and complex molecules (CM) defects) and for both diagnostic groups (Groups 1 and 2).

In conclusion, clinical assessments in combination with, consistent biomarkers are useful tools for increasing the diagnostic yield in IEM patients. In these cases, the use of targeted gene panels investigated by NGS is highly productive and cost-effective. The patients with no specific biomarkers represent more complexity. However, even if further clinical investigation is needed, the NGS panel approach yields important results and may help distinguish those patients who require further investigation with an additional exome/genome sequencing approach.

## Next-generation sequencing for diagnosis of rare diseases in the neonatal intensive care unit

TruSight One panel

Hussein Daoud PhD, Stephanie M. Luco BSc, Rui Li PhD, Eric Bareke PhD, Chandree Beaulieu MSc, Olga Jarinova PhD, Nancy Carson PhD, Sarah M. Nikkel MD, Gail E. Graham MD, Julie Richer MD, Christine Armour MSc MD, Dennis E. Bulman PhD, Pranesh Chakraborty MD, Michael Geraghty MD, Matthew A. Lines MD, Thierry Lacaze-Masmonteil PhD MD, Jacek Majewski PhD, Kym M. Boycott PhD MD, David A. Dyment DPhil MD

Integration of next-generation sequencing will enable molecular diagnosis during the hospital stay soon after birth, instead of families having to wait months to years for a diagnosis, which is the current norm.

**Table 3:** Details of mutations identified in patients with a positive molecular diagnosis

| Trio* | Sex | Affected gene | Inheritance           | Mutation type       | NCBI RefSeq    | cDNA and protein changes identified | Molecular diagnosis (OMIM no.)                                      |
|-------|-----|---------------|-----------------------|---------------------|----------------|-------------------------------------|---|
| 2     | M   | ACE           | Compound heterozygous | Frameshift deletion | NM_000789.3    | c.819_820delAG; p.(Arg274Glyfs*117) | Renal tubular dysgenesis (106180)                                   |
|       |     |               |                       | Frameshift deletion | NM_000789.3    | c.3521delG; p.(Gly1174Alafs*12)     |   |
| 6     | M   | SCN1A         | De novo               | Missense            | NM_001202435.1 | c.620T>G; p.(Val207Gly)             | SCN1A-related encephalopathy syndrome (607208)                      |
| 8     | M   | MTM1          | X-linked              | Nonsense            | NM_000252.2    | c.584C>A; p.(Tyr198*)               | Myotubular myopathy, X-linked (310400)                              |
| 9     | F   | FTO           | Autosomal recessive   | Missense            | NM_001080432.2 | c.956C>T; p.(Ser319Phe)             | FTO deficiency syndrome (612938)                                    |
| 11    | M   | WDR19         | Compound heterozygous | Nonsense            | NM_025132.3    | c.1600G>T; p.(Glu534*)              | Cranioectodermal dysplasia (614376)                                 |
|       |     |               |                       | Missense            | NM_025132.3    | c.2129T>C; p.(Leu710Ser)            |   |
| 15    | M   | CHRND         | Autosomal recessive   | Splice site         | NM_000751.2    | c.932+5G>A; p.?                     | Congenital myasthenic syndrome (601462)                             |
| 18    | M   | DYRK1A        | De novo               | Splice site         | NM_001396.3    | c.951+4_951+7delAGTA; p.?           | Autosomal dominant intellectual disability syndrome type 7 (614104) |
| 19    | F   | WT1           | De novo               | Missense            | NM_024426.4    | c.1460A>C; p.(His469Pro)            | Denys-Drash syndrome (194080)                                       |

Note: cDNA = complementary DNA, NCBI = National Center for Biotechnology Information (US), OMIM = Online Mendelian Inheritance in Man.

\*Trio = newborn + parents.

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# Adapt or retire?

- NGS services will be concentrated in centralised genetic centres. Choice of virtual panels will be directed by clinical input. Interpretation of results will be based on SIFT scores and clinical input.
- Confirmatory testing by metabolite or enzyme assay will not be sufficient to sustain small specialist laboratories. Expertise in specialist areas will be lost
- Routine services (eg AA or OA) with TAT in weeks will become irrelevant.